

Engineering Cas12j3 for Enhanced Trans-Cleavage Activity Toward Sensitive Nucleic Acid Detection

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Abstract

The trans-cleavage activity of CRISPR nucleases is critical for the development of highly sensitive nucleic acid detection platforms. Among compact effectors, Cas12j3 shows promising programmability but relatively limited trans-cleavage efficiency, which restricts its diagnostic potential. To address this limitation, we report a directed evolution strategy to enhance Cas12j3 activity using a two-stage screening workflow that integrates GFP-based primary screening and Cell-Free Protein Synthesis (CFPS)-based functional validation. In the first stage, a mutagenesis library of Cas12j3 variants was generated and expressed in a fluorescence-based screening system. A GFP-linked cleavage reporter enabled rapid identification of mutants with enhanced cis-cleavage activity. Variants showing increased GFP signal reduction were selected as candidates for further evaluation. In the second stage, these selected mutants were quantitatively assessed for trans-cleavage activity in a CFPS system. Fluorescence reporter assays were performed to compare the catalytic performance of evolved variants with the wild-type enzyme. This two-step strategy allows rapid enrichment of functionally improved variants while minimizing false positives due to expression variability. The evolved Cas12j3 variants exhibited significantly enhanced trans-cleavage activity, demonstrating the effectiveness of this approach. Overall, this platform provides a generalizable strategy for protein engineering of compact CRISPR nucleases and advances the development of future nucleic acid detection platforms with improved sensitivity and diagnostic potential.

Keywords

Cas12j3, Trans-Cleavage Activity, GFP-Based Screening, Nucleic Acid Detection